

Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ **DNA barcoding: case study #1**

Plant materials: landraces belonging to Mesoamerican and Andean gene pools

- 18 Central America
 - Mexico (13)
 - Guatemala (1)
 - Costa Rica (1)
 - El Salvador (1)
 - Honduras (1)
 - Venezuela (1)
- 12 Southern America
 - Argentina (6)
 - Bolivia (2)
 - Peru (1)
 - Cile (1)
 - Colombia (1)
 - Equador (1)

PLANT MATERIALS
List of 30 common bean landraces from the American gene pools

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Padua University

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ **DNA barcoding: case study #1**

Plant materials:
• 25 Italian varieties

- 1. Cannelino rosso (Pv)
- 2. Riso giallo (Pv)
- 3. Montalbano (Pv)
- 4. Munachedda nera (Pv)
- 5. San Michele (Pv)
- 6. Naseddu viola (Pv)
- 7. Maruchetta (Pv)
- 8. Riso bianco (Pv)
- 9. Cannelino nano (Pv)
- 10. Verdolino (Pv)
- 11. Blu lake (Pv)
- 12. Goldrush (Pv)
- 13. Cico (Pv)
- 14. BAT881 (Pv)
- 15. Lena (Pv)
- 16. Giulina (Pv)
- 17. Saluggia nano (Pv)
- 18. Venera (Pv)
- 19. Bofiolto Lampon (Pv)
- 20. Saluggia (Pv)
- 21. Cannelino (Pv)
- 22. Verdona (Pv)
- 23. San Matteo (Pv)
- 24. Zoffierino rovigotto (Pv)
- 25. Nero messicano (Pv)
- 26. F. di Lmia (Pi)
- 27. F. di Spagna (Pi)
- 28. F. dall'occhio (Vu)

• Accessions of *P. coccineus*, *P. lunatus* and *Vigna unguiculata* were adopted as out-types.

PLANT MATERIALS
List of 25 Italian common bean varieties

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ **Primers for bean DNA barcoding**

Design of primers to be used as chloroplast and nuclear barcodes.

TOOLS AND METHODS
How to design forward and reverse primers

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ **Primers for bean DNA barcoding**

List of primers used for chloroplast and nuclear markers.

Marker	Amplicon length (bp)			Primer name	Primer sequence (5'-3')
	<i>P. vulgaris</i>	<i>P. coccineus</i>	<i>P. lunatus</i>	<i>V. unguiculata</i>	
rbcL gene	543	543	543	rbcL_F	GACGATTCYCGAATRAAATCCYCA
				rbcL_R	GAAGGYTCTCTCAWCGGATAAA
matK gene	655	655	655	matK_724R*	TCACATGTACCTGCAGTAGC
				matK_1932a	CCTTCGATATCGGTTAAGAT
trnL intron	350	350	296	trnL_F	GGATAGTCCAGAGACTATATGGAAG
				trnL_R	TGACATGATTAATGGGACTATCTTTAT
ppb-rbcL ITS	329	325	325	ppb_F	GGAATCGGAGAGACTTAGC
				ppb_R	GGGATAGAGAGGACTTAGC
ppb-psbA ITS	365	365	365	ppb_F	GGTACTATCAATCACTCTCTTTAATGT
				ppb_R	ATCTAACTCTCGATATTAATAGCCAGC
ppb-trnT ITS	1117	1117	1124	ppb_F	GGCAACCAATCTTTGTTTC
				ppb_R	GATATGATGATGATGATGATGATG
trnT-trnL ITS	813	837	823	trnT_F	TTCTTTGAAATCCGATAGC
				trnT_R	CAAAATGATGCTTAACCT

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✓ **Bean DNA barcoding: experimental strategy**

1. Amplification by sequence-tagged site PCR
2. Sequencing of DNA amplicons
3. Multi-alignment of DNA sequences

STRATEGY
Experimental methods for DNA barcoding in common bean

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ **Bean DNA barcoding: results**

Basic information on the cpDNA and ITS barcode regions, including sequence length of amplicons, inter- and intra-specific number and frequency of SNPs and IN/DELS.

	rbcL	matK	trnL	ppb-rbcL	trnL-psbA	trnT-trnL	ppb-trnT	ITS1	ITS2
Total No. of <i>Phaseolus</i> primers	63	63	63	63	63	63	63	63	63
Average amplicon length (bp)	543	655	338	329	366	936	1124	358	413
No. SNPs in <i>Phaseolus</i> spp.	8	n.d.	21	14	14	53	48	65	58
Interspecific frequency (SNP/100 bp)	1.5	n.d.	6.0	4.3	3.8	6.5	4.2	17.4	13.8
No. SNPs in <i>P. vulgaris</i>	0	n.d.	4	0	0	9	2	6	4
Intraspecific frequency (SNP/100 bp)	0	n.d.	1.1	0	2.2	0.4	0.2	1.6	1.0
No. of In/Dels in <i>Phaseolus</i> spp.	0	n.d.	1	4	0	5	5	10	5
Average In/Del size (bp)	0	n.d.	58	2	0	7	2	4	5
No. of heterologous sites	n.a.	n.s.	n.s.	n.a.	n.s.	n.s.	n.s.	3	2
Amplification success (%)	100%	100%	100%	100%	100%	100%	100%	100%	100%
Sequencing success (%)	100%	82%	100%	100%	100%	100%	90%	97%	100%

n.d., not determined; n.a., not applicable.

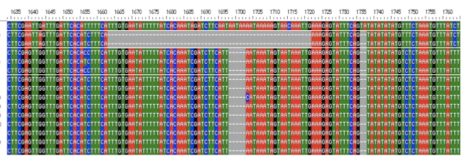
• The average frequency was 0.8 SNPs/100 bp (cpDNA) and 1.3 SNPs/100 bp (ITS) at the intraspecific level (*P. vulgaris*).

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results
Polymorphism information content of the investigated plastid genic regions



RESULTS
Finding SNPs and IN/DELS between *Vigna unguiculata* and *Phaseolus* species



- Several SNPs and IN/DELS were found between *Phaseolus* species and *Vigna unguiculata*

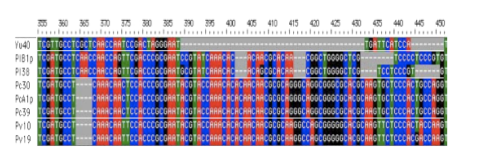
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results
IN/DELS and SNPs among *Phaseolus* spp. varieties



RESULTS
Finding SNPs and IN/DELS among *Phaseolus* species



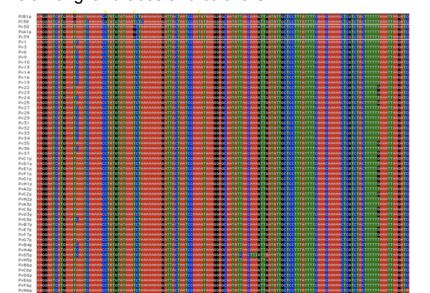
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results
SNPs among landraces and cultivars



RESULTS
Finding SNPs within *Phaseolus* species



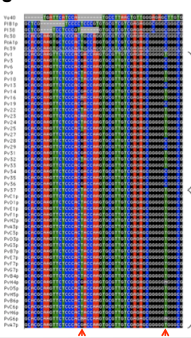
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results
SNPs among landraces and cultivars



RESULTS
Finding SNPs between *Phaseolus* gene pools and among landraces

- Italian cultivars
- Mesoamerican and Andean landraces



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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

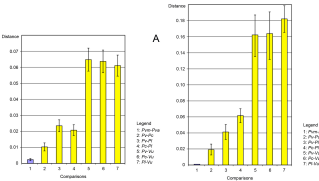
✓ Bean DNA barcoding: results
Mean and standard deviation of inter- and intra-specific genetic divergences calculated using the K2P distance model for the cpDNA markers and ITS regions, and overall.


Interspecific K2P distance	rbcL	trnT-trnL	trnK-trnD	trnL-trnF	trnL-trnC	trnT-trnL	trnK-trnD	trnL-trnF	trnL-trnC	Overall	St. Dev.	ITS	ITS2	Overall	St. Dev.
<i>P. vulgaris</i> / <i>P. coccineus</i>	0.0037	0.0139	0.0072	0.0107	0.0088	0.0070	0.0133	0.0020	0.0105	0.0169	0.0173	0.0065			
<i>P. vulgaris</i> / <i>P. sativus</i>	0.0074	0.0200	0.0094	0.0226	0.0227	0.0209	0.0214	0.0099	0.0080	0.0438	0.0432	0.0107			
<i>P. vulgaris</i> / <i>Vigna unguiculata</i>	0.0160	0.0460	0.0160	0.0380	0.0402	0.0371	0.0318	0.0276	0.0267	0.1871	0.1851	0.0231			
Interspecific K2P distance (<i>P. vulgaris</i>)	0.0000	0.0041	0.0001	0.0030	0.0008	0.0002	0.0013	0.0006	0.0002	0.0016	0.0006	0.0000	0.0000	0.0000	0.0003
St. Dev.	0.0000	0.0023	0.0001	0.0015	0.0005	0.0002	0.0009	0.0002	0.0002	0.0011	0.0005	0.0000	0.0000	0.0000	0.0003

RESULTS
The genetic divergence between *Phaseolus vulgaris* accessions was very low!

Genetic distances within and between *Phaseolus* spp. and *Vigna*:

inter- and intra-specific genetic distances referred to cpDNA markers (A) and ITS regions (B).





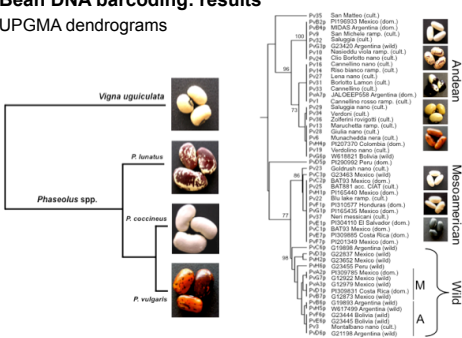
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results
UPGMA dendrograms



RESULTS
DNA barcoding is useful to identify *Phaseolus* species and to resolve diversity between bean species



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RESULTS
SNP-based haplotypes may discriminate Andean and Mesoamerican gene pools

1398 0 96763 96 0 6
91780927059401

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results

Consensus sequence related to the 17 individual SNPs detected in the target cpDNA regions with information on the haplotypes found across all entries.

Haplotype	No. Entries	trnL Intron			trnH-psaA							trnT-trnL		psb-trnC		
		G	A	T	A	T	A	A	G	T	C	A	A	T	G	A
Consensus sequence		G	A	T	A	T	A	A	G	T	C	A	A	T	G	A
Hap16	2	A	A		C	T	T	T	A							
Hap09	1				C	C	T	T	T	A					n.d.	
Hap03	10		C								G			G	C	
Hap08	1										G				C	
Hap12	1										G				C	
Hap13	3	A	G	A							G			G	C	
Hap06	7	A	C	A							G				C	
Hap14	1	A	C	A							G				C	
Hap15	3	A	C	A							G				C	
Hap02	16				C											
Hap04	3										G			T	T	
Hap10	1										G			T	T	
Hap11	1										G			T	T	
Hap07	1										G			T	T	
Hap05	6													n.d.		

n.d., not determined

■ Ancestrals
■ Mesoamerican gene pool
■ Andean gene pool

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RESULTS
Genetic diversity is about 25% higher in the Mesoamerican than in the Andean gene pool

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results

Summary of haplotype statistics computed separately for chloroplast (A) and nuclear (B) DNA markers.

A

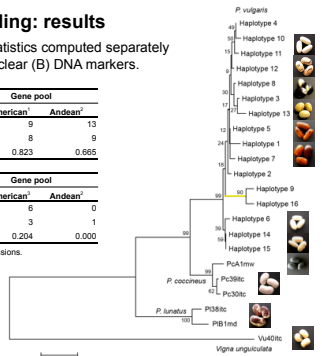
Genetic diversity statistics	Gene pool	
	Mesoamerican ¹	Andean ²
No. segregating sites (S)	9	13
Haplotype number (Hn)	8	9
Haplotype diversity (Hd)	0.823	0.665

B

Genetic diversity statistics	Gene pool	
	Mesoamerican ¹	Andean ²
No. segregating sites (S)	6	0
Haplotype number (Hn)	3	1
Haplotype diversity (Hd)	0.204	0.000

¹29 accessions; ²26 accessions; ³26 accessions.

NJ tree based on the haplotypes identified for *Phaseolus* accessions.



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RESULTS
The population structure analyses revealed 3 distinct subgroups

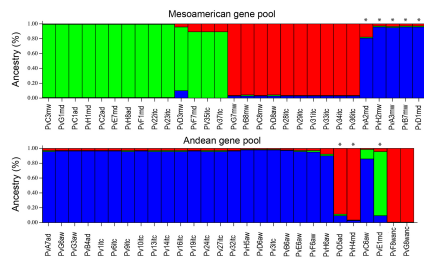
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results

Population structure of *Phaseolus vulgaris* germplasm core collection as estimated with STRUCTURE software



• Each accession is represented by a vertical histogram partitioned into K=3 colored segments that represent the estimated membership of each individual.

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RESULTS
Most Italian varieties (i.e. pure lines) are genetically differentiated from American landraces

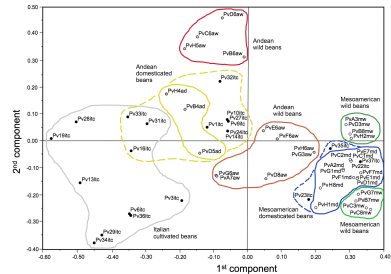
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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: results

Genetic diversity and differentiation of *P. vulgaris* accessions



• Italian varieties originated from both Andean and Mesoamerican gene pools.

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CASE STUDY
Concluding remarks on DNA barcoding in common beans

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: concluding remarks

- Strategy: multi-locus vs. single-locus barcoding
 - The occurrence of SNPs was lower in cpDNA barcodes than ITS-1-2 regions (0.8 vs. 1.3 per 100 bp);
- Strategy: NJ tree-based vs. Haplotype-based
 - The NJ method is a powerful approach for estimating genetic differentiation among species, but it is relatively uninformative for studying genetic diversity within species;
 - The character-based method is able to identify several distinct haplotypes over all target regions corresponding to Mesoamerican or Andean accessions.
- Most domesticated Andean varieties were clustered apart from Mesoamerican varieties;
 - The Italian varieties originated from both Andean (15) and Mesoamerican (10) gene pools.

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CASE STUDY
Use of cpDNA barcoding in common bean

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Plastidial DNA barcoding for assessing gene pool diversity in common bean (*Phaseolus* spp.)

✓ Bean DNA barcoding: concluding remarks

- The DNA barcoding based on plastid genic and intergenic sequences confirmed to be a very powerful tool for reconstructing genetic relationships among plant species.
- Our findings raise some concerns about the use of plastid DNA barcoding for intraspecific genetic diversity studies in plant species and highlights its limitations for resolving genetic relationships between landraces and cultivars.



Case study #2: Nuclear DNA barcoding for assessing genetic identity of grapevine cultivars

✓ Identification of grapevine cultivars
DNA barcoding as a tool for testing genetic identity within species!

BACKGROUND
Identification of grapevine cultivars (i.e. below the species level)

Ampelography
→ leaves, shoots, flowers, bunches, berries, etc.
Protein/Metabolite profiling
→ proteins, secondary metabolites (anthocyanins)
DNA genotyping
→ SSR markers (Vitis Intl. Catalogue of Cultivars)

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Multilocus DNA barcoding of grapevine germplasm

- Implementation of a molecular diagnostic assay for the genetic identification of grapevine cultivars, grapes and wines through the reconstruction of cultivar-specific haplotypes;

GOALS
Multilocus DNA barcoding in *Vitis vinifera* L.

- Resolution of homonym/synonymy of grapevine cultivars;
- Traceability of grapevine-derived products (grapes and wines).

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Plant materials: *Vitis* spp. accessions
A total of 152 accessions of *Vitis vinifera* L. were analyzed for the discovery of single-copy gene-specific SNPs

PLANT MATERIALS
152 accessions of *Vitis vinifera* L. and 5 wild species as outgroups

- V. labrusca*
- V. riparia*
- V. berlandieri*
- V. rupestris*
- V. cinerea*
- V. vinifera* L.

- 5 outgroups
- 127 European certified cultivars
- 25 Italian local cultivars

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Selection of barcode regions
Common target genes for DNA barcoding

Gene*	Genomic location	Animals	Plants	Fungi
COI-barcode [†]	Mitochondria	196 777	520	1 931
ITS-DNA	Mitochondria	41 361	221	2 058
cyb	Mitochondria	89 324	165	1 920
ITS1-rDNA	Nucleus	12 176	57 663	66 269
ITS2-rDNA	Nucleus	13 923	58 065	67 332
ITS-DNA	Nucleus	21 063	17 121	32 280
rbcL	Plastid	N/A	20 063	37 328

TOOLS AND METHODS
Selection of the target sequences for DNA barcoding

Marker	Chromosome	Gene product	Length (bp)	Primer name	Primer sequence (5'-3')	Ta (°C)	References
GAJ [‡]	1	Transcription factor for gibberellins	761	GAJ_F	ATGGATGAGCTCTCGCTGT	50	Wien et al. (2007)
IDOL [§]	3	Predicted ZIP-DNA-binding protein	419	IDOL_F	CACCACTCCCTACCACTGTG	55	Salmasso et al. (2004)
ICOL [¶]	3	Predicted zinc-finger protein	418	ICOL_F	CAAGGCTCTCTCTGATAC	60	Salmasso et al. (2004)
ATPS	7	ATP synthase	800	ATP_F	ATGCTTCCAGTCCGCTTC	60	Nicolò et al. (unpublished)
UFGT [‡]	16	UDP-glucose flavonoid 3-O-glucosyltransferase	910	UFGT_F4	ATGCTCAAGCAACCAACAC	63	Nicolò et al. (unpublished)

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Strategy adopted for SNP discovery and CV haplotyping

STRATEGY
Experimental strategy adopted for DNA barcoding

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Discovery of discriminant SNPs
SNPs among *Vitis* species

SNPs among *V. vinifera* cultivars: 1 SNP/34 bp (23-51)

SNPs among biotypes within *V. vinifera* cultivars

RESULTS
Discovery of discriminant SNPs either between *Vitis* species or within *V. vinifera* species

Heterozygous SNPs in *V. vinifera* cultivars

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RESULTS
Genetic diversity statistics were high for Italian cultivars whereas low genetic differentiation was found among cultivar subgroups (18%)

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Gene diversity descriptive statistics

Overall number of polymorphic sites, observed and expected number of nucleotides per SNP site, information index of phenotypic diversity (SI) and polymorphism information content (PIC)

	No. A	P (%)	na	ne	SI	PIC
Italy	164	68 (62.4%)	1.67	1.19	0.201	0.140
Greece	22	39 (35.8%)	1.37	1.22	0.187	0.125
Portugal	32	41 (37.6%)	1.39	1.19	0.180	0.116
Rumania	8	33 (30.3%)	1.31	1.19	0.168	0.112
Spain	36	43 (39.4%)	1.42	1.23	0.195	0.128
Local	44	44 (40.4%)	1.42	1.17	0.169	0.106
Hybrid	12	56 (51.4%)	1.52	1.32	0.285	0.190
Outgroup	10	67 (61.5%)	1.74	1.40	0.361	0.123
Cultivars	274	93 (85.3%)	1.94	1.21	0.219	0.134
Outgroups	10	67 (61.5%)	1.74	1.40	0.361	0.123
Total	313	108 (99.1%)	2.22	1.21	0.240	0.140
St. Dev.	14	13 (0.0%)	0.50	0.28	0.220	0.160

Summary of F-statistics and gene flow (Nm) estimates over all SNPs and accessions.

	n	F-statistics			
		Fis	Fit	Fst	Nm
Total	313	-0.09	0.10	0.18	1.12

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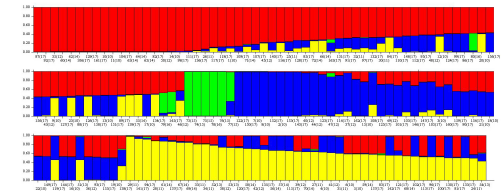
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RESULTS
A total of 4 distinct genetic clusters inferred out of the 8 sample subgroups predefined according to the geographical origin of cultivars

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Population structure analysis (STRUCTURE v2.1)



Admixed ancestry: 54% of accessions (q < 0.7)

Subgroup	Inferred clusters				No. of individuals
	1	2	3	4	
Local	0.47	0.00	0.37	0.16	23
Italy	0.47	0.01	0.33	0.19	83
Greece	0.38	0.01	0.14	0.50	11
Portugal	0.35	0.01	0.42	0.22	16
Rumania	0.21	0.01	0.18	0.60	4
Spain	0.51	0.01	0.07	0.42	18
Hybrid	0.35	0.53	0.03	0.01	3
Outgroup	0.01	0.52	0.07	0.01	5

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RESULTS
Construction of NJ trees based on the coalescence of specific populations

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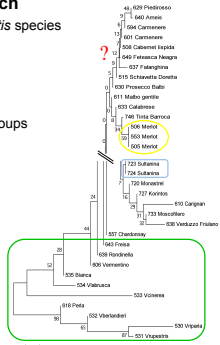
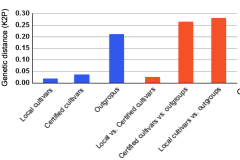
Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Phenetic tree-building approach

NJ tree showing relationships among *Vitis* species and *V. vinifera* cultivars

Genetic distances

within (■) and between (■) accessions groups



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RESULTS
Summary of SNP statistics for the nuclear single-copy barcode genes: all variable SNP sites were used (pi > 95%)

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Character-based approach

detection of specific informative character states, SNPs or In/Dels, shared by taxonomic subgroups (biotypes and cultivars) and suitable for the identification of diagnostic haplotypes

- 1: AAACCTT
- 2: AAACTT
- 3: AAACCTT

- 1 SNP/34 bp
- Hn=127/152 CVs

Gene	Length (bp)	No. SNPs		Frequency (bp/1 SNP)		Hn
		<i>Vitis</i> spp.	<i>V. vinifera</i>	<i>Vitis</i> spp.	<i>V. vinifera</i>	
GAI	761	18	15	42.27	53.73	23
ID04	419	21	20	19.95	20.95	33
IC08	418	20	18	20.50	23.22	14
ATP	800	21	15	38.09	53.33	25
UFGT	919	29*	29*	31.69	31.69	97
Combined	3317	109	97	30.43	34.19	132

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RESULTS
Multi-locus haplotype composition: heterozygous sites were dealt as functionally haploid SNPs (using IUB codes)

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Multi-locus haplotypes

(3 nuclear barcode regions)

Overall:
53 SNPs over 1,598 bp
(1 SNP/29 bp)
62 haplotypes including 1 (38) to 9 accessions

- Replicated accessions:
- Sultanina
 - Carmener
 - Malbec
 - Cannonao
 - Sagrantino
 - Raboso piave

- Synonymies:
- Merlot (cultivars vs. locals)
 - Raboso piave vs. Friularo
 - Marzemina (nera vs. cener).

Genotype/accession	Hp	GAI	ID04	IC08
PI11_P11merlot	1
PI11_P11carmener	1
PI11_P11malbec	1
PI11_P11cannonao	1
PI11_P11sagrantino	1
PI11_P11rabosopiave	1
PI11_P11merlot	2
PI11_P11carmener	2
PI11_P11malbec	2
PI11_P11cannonao	2
PI11_P11sagrantino	2
PI11_P11rabosopiave	2
PI11_P11merlot	3
PI11_P11carmener	3
PI11_P11malbec	3
PI11_P11cannonao	3
PI11_P11sagrantino	3
PI11_P11rabosopiave	3
PI11_P11merlot	4
PI11_P11carmener	4
PI11_P11malbec	4
PI11_P11cannonao	4
PI11_P11sagrantino	4
PI11_P11rabosopiave	4
PI11_P11merlot	5
PI11_P11carmener	5
PI11_P11malbec	5
PI11_P11cannonao	5
PI11_P11sagrantino	5
PI11_P11rabosopiave	5
PI11_P11merlot	6
PI11_P11carmener	6
PI11_P11malbec	6
PI11_P11cannonao	6
PI11_P11sagrantino	6
PI11_P11rabosopiave	6
PI11_P11merlot	7
PI11_P11carmener	7
PI11_P11malbec	7
PI11_P11cannonao	7
PI11_P11sagrantino	7
PI11_P11rabosopiave	7
PI11_P11merlot	8
PI11_P11carmener	8
PI11_P11malbec	8
PI11_P11cannonao	8
PI11_P11sagrantino	8
PI11_P11rabosopiave	8
PI11_P11merlot	9
PI11_P11carmener	9
PI11_P11malbec	9
PI11_P11cannonao	9
PI11_P11sagrantino	9
PI11_P11rabosopiave	9

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RESULTS
Single-copy gene SNPs and haplotype composition

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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Single-locus haplotypes

UFGT as barcode region: 919 bp (1 SNP/32 bp)



92 haplotypes out of 152 cultivars including 1 to 12 accessions each

Cultivars/haplotype #

- 1 H : 1 CV 68
- 1 H : 2 CVs 13
- 1 H : 3 CVs 6
- 1 H : 4 CVs 1
- 1 H : ≥5 CVs 4

Genotype/accession	Hp	SNP sites in the UFGT gene
PI11_P11merlot	1
PI11_P11carmener	1
PI11_P11malbec	1
PI11_P11cannonao	1
PI11_P11sagrantino	1
PI11_P11rabosopiave	1
PI11_P11merlot	2
PI11_P11carmener	2
PI11_P11malbec	2
PI11_P11cannonao	2
PI11_P11sagrantino	2
PI11_P11rabosopiave	2
PI11_P11merlot	3
PI11_P11carmener	3
PI11_P11malbec	3
PI11_P11cannonao	3
PI11_P11sagrantino	3
PI11_P11rabosopiave	3
PI11_P11merlot	4
PI11_P11carmener	4
PI11_P11malbec	4
PI11_P11cannonao	4
PI11_P11sagrantino	4
PI11_P11rabosopiave	4
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PI11_P11merlot	9
PI11_P11carmener	9
PI11_P11malbec	9
PI11_P11cannonao	9
PI11_P11sagrantino	9
PI11_P11rabosopiave	9

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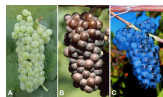
Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Single-locus haplotypes
UFGT as barcode region: 919 bp
(1 SNP/32 bp)



Table listing grapevine cultivars and their corresponding UFGT haplotypes. Columns include cultivar names (e.g., P133_Costabona, P133_Zanone) and their haplotypes (e.g., ...G...G...G...).

92 haplotypes out of 152 cultivars including 1 to 12 accessions each



RESULTS
Single-copy gene
SNP haplotype
composition



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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Single-locus haplotypes
UFGT as barcode region: 919 bp
(1 SNP/32 bp)



Table listing grapevine cultivars and their corresponding UFGT haplotypes. Columns include cultivar names (e.g., P133_Costabona, P133_Zanone) and their haplotypes (e.g., ...G...G...G...).

92 haplotypes out of 152 cultivars including 1 to 12 accessions each



RESULTS
Single-copy gene
SNP haplotype
composition



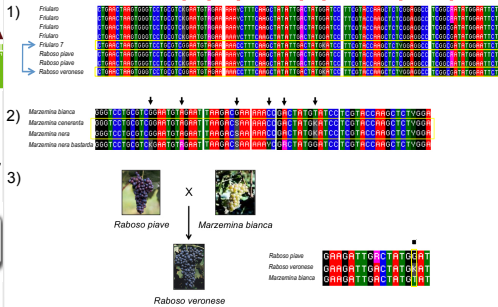
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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

✓ Specific findings
(mislabeling, synonymy/homonymy, hybrids)



FINDINGS
Grapevine DNA
barcoding revealed
cases of mislabeling,
synonymy and
hybridity



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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

Concluding remarks

- ✓ The chloroplast genome proved to be not enough polymorphic within *V. vinifera* species to distinguish grapevine cultivars;
- ✓ A multi-locus DNA barcoding based on five nuclear regions allowed to reconstruct cultivar-specific haplotypes (diagnostic characters fixed in single cultivars or cultivar subgroups):
 - many cultivar-specific haplotypes were found;
 - accessions belonging to a given cultivar clustered together sharing the same haplotype;
 - several cases of homonymy/synonymy were resolved;
 - the hybrid origin of some hybrids was verified;
 - local cultivars were classified correctly with the international certified cultivars.
- ✓ The single-copy gene UFGT is an excellent candidate for the nuclear DNA barcoding of grapevine cultivars.



CONCLUSIONS
Nuclear DNA
barcoding opens new
perspectives on the
genetic traceability of
grapevine cultivars



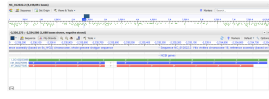
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Nuclear DNA barcoding for assessing the genetic identity of grapevine cultivars

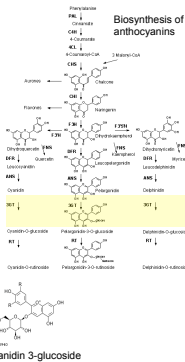
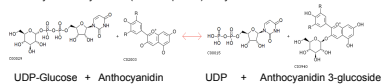
✓ Information on VvUFGT
Gene structure (1,536 nt, CDS=1,371 nt)



Protein (456 aa) domains and active sites



Biochemical function (glycosyl-transferase)
Anthocyanin biosynthesis (KEGG map00942) Enzyme 2.4.1.115



UFGT
The best candidate
gene for grapevine
DNA barcoding



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DNA barcoding and its potentials for assessing genetic identity in plant species

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